

Note

The synthesis of the 1-*N*-(L-aspart-4-oyl)glucosylamine linkage. New synthesis of 1-*N*-(L-aspart-1- and 4-oyl)-4-*O*- β -D-galactopyranosyl- β -D-glucopyranosylamines*

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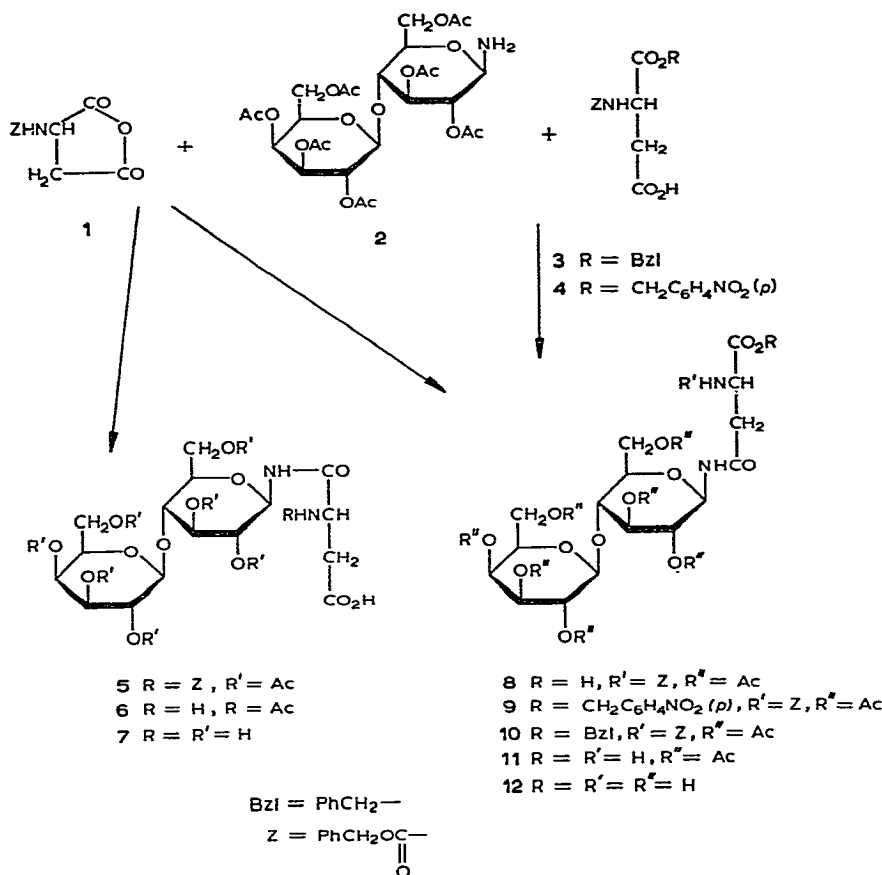
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Synthetic oligosaccharide-asparagine compounds are of interest for determining the specificity of lectins for which the asparagine residue may be a part of the immunodeterminant¹, and for determining the chemical structure of antigenic sites containing a 1-*N*-(L-aspart-4-oyl)glucosylamine component².

The methods of synthesis of 2-acetamido-1-*N*-(L-aspart-4-oyl)-2-deoxy- β -D-glucopyranosylamine, the glucosylamine acid residue found as the carbohydrate-protein linkage of many glycoproteins of excretory origin³, include both the reaction of the glucosylamine with *N*-(benzyloxycarbonyl)-L-aspartic anhydride⁴ (**1**) and with a partially protected L-aspartoyl derivative, 1-benzyl *N*-(benzyloxycarbonyl)-L-aspartate⁵ (**3**). In the first reaction, both the L-aspart-1- and 4-oyl derivatives are obtained and, consequently, yields lower than 50% are expected for each derivative. The possibility exists, however, that the reaction shows some specificity, which may vary with the type of reagent used⁶. One advantage of the first method over the second is that it avoids the preparation of the derivative protected at C-1, which may be both lengthy and give poor yields when applied to long-chain peptides. In order to test the stereoselectivity and the yield of the first method with an oligosaccharide, *N*-(benzyloxycarbonyl)-L-aspartic anhydride⁷ (**1**) was condensed with 2,3,6-tri-*O*-acetyl-4-*O*-(2,3,4,6-tetra-*O*-acetyl- β -D-galactopyranosyl)- β -D-glucopyranosylamine⁸ (**2**) to give the L-aspart-1- and 4-oyl compounds (**5** and **8**, respectively), which were separated by column chromatography.

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Both compounds were obtained in approximately the same yield and no stereoselectivity was observed. The total yield, based on the carbohydrate starting material, was low (33%), but the peptide starting material may be recovered, which is advantageous for peptides requiring a lengthy synthesis. Thus, the method may be used for the condensation of glycosylamines of readily obtainable disaccharides with peptides bearing an L-aspartoyl residue as C-terminal.

On hydrogenolysis in the presence of 10% palladium-on-charcoal, 2,3,6-tri-*O*-acetyl-1-*N*-[*N*-(benzyloxycarbonyl)-L-aspart-4-oyl]-4-*O*-(2,3,4,6-tetra-*O*-acetyl-β-D-galactopyranosyl)-β-D-glucopyranosylamine (**8**) gave **11**, which on treatment with methanolic ammonia afforded 1-*N*-(L-aspart-4-oyl)-4-*O*-(β-D-galactopyranosyl)-β-D-glucopyranosylamine (**12**), prepared earlier from the corresponding benzyl ester derivative^{8,9} (**10**). Similarly, 2,3,6-tri-*O*-acetyl-1-*N*-[*N*-(benzyloxycarbonyl)-L-aspart-1-oyl]-4-*O*-(2,3,4,6-tetra-*O*-acetyl-β-D-galactopyranosyl)-β-D-glucopyranosylamine (**5**) gave, on hydrogenation, 2,3,6-tri-*O*-acetyl-1-*N*-[L-aspart-1-oyl]-4-*O*-(2,3,4,6-tetra-*O*-acetyl-β-D-galactopyranosyl)-β-D-glucopyranosylamine (**6**), which was treated with

methanolic ammonia to afford 1-*N*-(L-aspart-1-oyl)-4-*O*-(β -D-galactopyranosyl)- β -D-glucopyranosylamine (7).

In the second type of synthesis of the 1-*N*-(L-aspart-4-oyl)glycosylamine linkage⁵, the 1-benzyl ester, which is used as a protecting group, has the disadvantage of being labile under the acidic conditions^{10,11} required for removal of the *N*-(benzyloxycarbonyl) protecting group, thus limiting the usefulness of this derivative for the elongation of the chain toward the N-terminal. In contrast, the *p*-nitrobenzyl ester exhibits a marked stability to acid cleavage¹², which permits selective removal of the *N*-(benzyloxycarbonyl) group. Therefore, the condensation of *N*-(benzyloxycarbonyl)-L-aspartic *p*-nitrobenzyl ester¹³ with 2 in the presence of 2-ethoxy-*N*-ethoxycarbonyl-1,2-dihydroquinoline¹⁴ (E.E.D.Q.) at room temperature gave pure 2,3,6-tri-*O*-acetyl-1-*N*-[1-*p*-nitrobenzyl *N*-(benzyloxycarbonyl)-L-aspart-4-oyl]-4-*O*-(2,3,4,6-tetra-*O*-acetyl- β -D-galactopyranosyl)- β -D-glucopyranosylamine (9) in almost quantitative yield, without requiring purification by chromatography. Hydrogenolysis of 9 in the presence of palladium-on-charcoal gave the partially protected derivative⁸ 11.

EXPERIMENTAL

General methods. — Melting points were determined with a Mettler FP-2 apparatus and correspond to "corrected melting points". Rotations were determined for solutions in 1-dm, semimicro tubes with a Perkin-Elmer No. 141 polarimeter. The *N,N*-dimethylformamide used was spectro-reagent grade. I.r. spectra were recorded for potassium bromide discs, with a Perkin-Elmer spectrophotometer Model 237. Evaporations were performed *in vacuo*, the bath temperature being kept below 45°. Column chromatography was performed on Silica Gel Merck (70–325 mesh, E. Merck, Darmstadt, Germany), used without pretreatment; the ratio of the weight of the substance to the weight of silica gel was 1:50; the volume of the fractions collected was 4 ml per g of the substance; and the ratio of diameter of the column to its length was 1:16. The homogeneity of compounds was verified by ascending t.l.c. on precoated plates of Silica Gel (Merck); the spots were detected by spraying with 20% sulfuric acid and heating for a few min at 200°. Spots of free amino glycopeptides were detected by spraying with a ninhydrin solution in acetone and heating the plates for a few seconds. Microanalyses were performed by Dr. W. Manser, Zurich, Switzerland.

2,3,6-Tri-*O*-acetyl-1-*N*-[*N*-(benzyloxycarbonyl)-L-aspart-1-oyl]-4-*O*-(2,3,4,6-tetra-*O*-acetyl- β -D-galactopyranosyl)- β -D-glucopyranosylamine (5) and 2,3,6-tri-*O*-acetyl-1-*N*-[*N*-(benzyloxycarbonyl)-L-aspart-4-oyl]-4-*O*-(2,3,4,6-tetra-*O*-acetyl- β -D-galactopyranosyl)- β -D-glucopyranosylamine (8). — A solution of *N*-(benzyloxycarbonyl)-L-aspartic anhydride⁷ (1, 0.45 g, calc. on the basis of a 50% yield of 2) in ethyl acetate (5 ml) was added to a solution of 2,3,6-tri-*O*-acetyl-4-*O*-(2,3,4,6-tetra-*O*-acetyl- β -D-galactopyranosyl)- β -D-glucopyranosylamine [2, obtained⁸ from 2.0 g of 2,3,6-tri-*O*-acetyl-4-*O*-(2,3,4,6-tetra-*O*-acetyl- β -D-galactopyranosyl)- β -D-glucopyranosyl azide] in ethyl acetate (10 ml). The solution was kept for 24 h at room temperature. The ethyl acetate layer was washed with M hydrochloric acid and water,

and dried (sodium sulfate). Removal of the solvent gave 1.75 g of product (93% yield, based on **1**) showing on t.l.c. in 19:1 (v/v) chloroform-methanol two spots, for **5** (R_F 0.4) and **8** (R_F 0.15), in addition to three spots having R_F 0.7, 0.9, and 0.95, corresponding to sugar compounds devoid of aspartoyl residue. A solution of the mixture (1.0 g) in chloroform was applied to a column of silica gel. Successive elutions with a linear gradient (1:0 to 10:1 v/v) of chloroform-methanol, gave at first 0.45 g containing only sugar residues, and then **5** (0.18 g, 18%), which was crystallized from methanol; m.p. 234.5–235.5°, $[\alpha]_D^{23} +20^\circ$ (c 1.2, chloroform); ν_{\max}^{KBr} 3350 (NH), 1750 (OAc), 1650 (shoulder, benzyloxycarbonyl CO), and 1550–1650 cm^{-1} (peptide Amide I); t.l.c. 9:1 (v/v) chloroform-methanol: R_F 0.37.

Anal. Calc. for $\text{C}_{38}\text{H}_{48}\text{N}_2\text{O}_{22}$: C, 51.58; H, 5.47; N, 3.17; O, 39.77. Found: C, 51.50; H, 5.40; N, 3.11; O, 39.61.

Compound **8** (0.15 g, 15%) was eluted after complete elution of **5**, and was crystallized from abs. ethanol; m.p. 177–178° (shrinks at 175°), $[\alpha]_D^{23} -6.7^\circ$ (c 1.2, chloroform) ν_{\max}^{KBr} 3400 (NH), 1760 (OAc), 1650 (shoulder, benzyloxycarbonyl CO), and 1525–1600 cm^{-1} (peptide Amide I); t.l.c. (9:1, v/v, chloroform-methanol): R_F 0.21.

Anal. Calc. for $\text{C}_{38}\text{H}_{48}\text{N}_2\text{O}_{22}$: C, 51.58 H, 5.47 N, 3.17 O, 39.77. Found: C, 51.65; H, 5.49; N, 3.15; O, 39.23.

2,3,6-Tri-O-acetyl-1-N-[1-p-nitrobenzyl N-(benzyloxycarbonyl)-L-aspart-4-oyl]-4-O-(2,3,4,6-tetra-O-acetyl- β -D-galactopyranosyl)- β -D-glucopyranosylamine (9). — To a solution of **2** (1 g) in benzene-ethanol (1:1, v/v, 20 ml) was added 1-p-nitrobenzyl *N*-(benzyloxycarbonyl)-L-aspartate¹³ (**4**, 0.63 g) and 2-ethoxy-*N*-ethoxycarbonyl-1,2-dihydroquinoline¹⁴ (0.4 g). The mixture was stirred overnight at room temperature, the solvents were evaporated, and the residue was dissolved in chloroform. The organic layer was washed successively with M hydrochloric acid, water, 1% sodium hydrogencarbonate, and water, dried (sodium sulfate), and evaporated. The residue crystallized from 1:1 (v/v) benzene-alcohol as needles (1.0 g, 98%), m.p. 113–114°, $[\alpha]_D^{22} +13^\circ$ (c 1.3, chloroform); ν_{\max}^{KBr} 3350 (NH) 1750 (OAc), 1610 (benzyloxycarbonyl CO), and 1525 cm^{-1} (peptide Amide I).

Anal. Calc. for $\text{C}_{45}\text{H}_{53}\text{N}_3\text{O}_{24}$: C, 52.98; H, 5.25; N, 4.13; O, 37.73. Found: C, 52.64; H, 5.20; N, 4.13; O, 38.12.

2,3,6-Tri-O-acetyl-1-N-(L-aspart-4-oyl)-4-O-(2,3,4,6-tetra-O-acetyl- β -D-glucopyranosyl)- β -D-glucopyranosylamine (11). — *A.* A solution of **9** (0.5 g) in 10:1 (v/v) methanol-water (110 ml) was hydrogenated in the presence of 10% palladium-on-charcoal (0.15 g) for 3 h at room temperature and at a pressure of 2.4 atm. The catalyst was filtered off, and the filtrate evaporated to dryness to give **11** (0.35 g, 93%), m.p. 196–197°, $[\alpha]_D^{22} -6.8^\circ$ (c 0.91, chloroform); ν_{\max}^{KBr} 1750 (OAc) cm^{-1} ; t.l.c. (4:1:5, v/v, butanol-acetic acid-water): R_F 0.32 (brown color with ninhydrin reagent).

Anal. Calc. for $\text{C}_{30}\text{H}_{42}\text{N}_2\text{O}_{20}$: C, 48.00; H, 5.64; N, 3.73; O, 42.62. Found: C, 48.08; H, 5.66; N, 3.80; O, 42.50.

B. A solution of **8** (0.08 g) in 6:1 (v/v) ethanol-acetic acid (35 ml) was hydrogenated in the presence of 10% palladium-on-charcoal (0.05 g) for 3 h at room

temperature. The catalyst was filtered off and the filtrate was evaporated. The residue crystallized from hot abs. ethanol to give 40 mg (58%), m.p. 197–198; i.r. spectra and t.l.c. mobility were identical with those of the compound obtained from method A.

1-N-(L-Aspart-4-oyl)-4-O-β-D-galactopyranosyl-β-D-glucopyranosylamine (12). — A solution of crude 11 (obtained from 0.5 g of 9) in dry methanol (15 ml) was saturated with ammonia at 0° and kept overnight. The solvent was evaporated *in vacuo* at room temperature. The residue was washed with acetone to remove acetamide and crystallized from water-methanol to give 12 (0.11 g, 52%), m.p. 235–235° (dec.), $[\alpha]_D^{22} +1.8^\circ$ (c 1.1, water); ν_{\max}^{KBr} 3350 (broad, OH) and 1550–1675 cm^{-1} (peptide Amide I); t.l.c. (4:1, v/v pyridine–water): R_F 0.12 (brown color with ninhydrin); lit.⁹: m.p. 235° (dec.), $[\alpha]_D^{22} +1.0^\circ$ (water).

Anal. Calc. for $\text{C}_{16}\text{H}_{28}\text{N}_2\text{O}_{13}\cdot\text{H}_2\text{O}$: C, 40.60; H, 6.37; N, 5.90; O, 47.21. Found: C, 40.38; H, 6.44; N, 5.90; O, 47.40.

2,3,6-Tri-O-acetyl-1-N-(L-aspart-1-oyl)-4-O-(3,2,4,6-tetra-O-acetyl-β-D-glucopyranosyl)-β-D-glucopyranosylamine (6). — A solution of 5 (0.35 g) in 10:1 (v/v) ethanol–acetic acid was hydrogenated in the presence of 10% palladium-on-charcoal for 3 h at room temperature and at a pressure of 2.0 atm. The catalyst was filtered off, the solution evaporated, and the residue crystallized from hot abs. ethanol to give 6 (0.27 g, 95%), m.p. 154–155° (dec.), $[\alpha]_D^{22} +3.8^\circ$ (c 1.2, chloroform); ν_{\max}^{KBr} 1750 (OAc) and 1575 cm^{-1} (peptide Amide I); t.l.c. (4:1:5, v/v, butanol–acetic acid–water): R_F 0.30 (purple color with ninhydrin reagent).

Anal. Calc. for $\text{C}_{30}\text{H}_{42}\text{N}_2\text{O}_{20}$: C, 48.00; H, 5.64; N, 3.73; O, 42.62. Found: C, 47.94; H, 5.68; N, 3.81; O, 42.69.

1-N-(L-Aspart-1-oyl)-4-O-β-D-galactopyranosyl-β-D-glucopyranosylamine (7). — A solution of crude 6 (obtained from 0.25 g of 5) in dry methanol (20 ml) was saturated with ammonia at 0°, and kept overnight at 0°. The solution was concentrated *in vacuo* at room temperature. The resultant crystalline material was filtered off, washed with acetone (75 mg), and recrystallized from water-methanol, giving 7 (50 mg, 48%), m.p. 218–220° (dec.) (browning at 196°), $[\alpha]_D^{22} +14^\circ$ (c 0.79, water); ν_{\max}^{KBr} 3350 (broad, OH) and 1550–1680 cm^{-1} (peptide Amide I); t.l.c. (4:1, v/v pyridine–water): R_F 0.16 (purple color with ninhydrin).

Anal. Calc. for $\text{C}_{16}\text{H}_{28}\text{N}_2\text{O}_{13}\cdot\text{H}_2\text{O}$: C, 40.60; H, 6.37; N, 5.90; O, 47.21. Found: C, 40.39; H, 6.44; N, 5.90; O, 47.40.

REFERENCES

- 1 S. TOYOSHIMA, T. AKIYAMA, K. NAKANO, A. TONOMURA, AND T. OSAWA, *Biochemistry*, 10 (1971) 4457–4463; S. TOYOSHIMA, M. FUKUDA, AND T. OSAWA, *ibid.*, 11 (1972) 4000–4005; T. KAWAGUCHI, I. MATSUMOTO, AND T. OSAWA, *ibid.*, 13 (1974) 3169–3173; R. KAIFU, T. OSAWA, AND R. W. JEANLOZ, *Carbohydr. Res.*, 40 (1975) 111–117; R. KORNFIELD AND S. KORNFIELD, *J. Biol. Chem.*, 245 (1970) 2536–2545; S. KORNFIELD, J. ROGERS, AND W. GREGORY, *ibid.*, 246 (1971) 6581–6586.
- 2 C. BANJO, P. GOLD, C. W. GEHRKE, S. O. FREEDMAN, AND J. KRUPPEY, *Int. J. Cancer*, 13 (1974) 151–163; R. VRBA, E. ALPERT, K. J. ISSELBACHER, AND R. W. JEANLOZ, *Immunochemistry*, in press.
- 3 R. D. MARSHALL AND A. NEUBERGER, in A. GOTTSCHALK (Ed.), *Glycoproteins*, 2nd edn., Elsevier, Amsterdam, 1972, pp. 453–470.
- 4 H. G. GARG AND R. W. JEANLOZ, *Carbohydr. Res.*, 23 (1972) 437–439.

- 5 G. S. MARKS, R. D. MARSHALL, AND A. NEUBERGER, *Biochem. J.*, 87 (1963) 274-281; C. H. BOLTON AND R. W. JEANLOZ, *J. Org. Chem.*, 28 (1963) 3228-3230.
- 6 W. J. LEQUESNE AND G. T. YOUNG, *J. Chem. Soc.*, (1952) 24-28.
- 7 Y. YAMAMOTO, *Biochem. Prep.*, 10 (1953) 10-18.
- 8 M. SPINOLA AND R. W. JEANLOZ, *Carbohydr. Res.*, 15 (1970) 361-369.
- 9 D. DUNSTAN AND L. HOUGH, *Carbohydr. Res.*, 23 (1972) 17-21.
- 10 D. BEN-ISHAI AND A. BERGER, *J. Org. Chem.*, 17 (1952) 1564-1570; D. BEN-ISHAI, *ibid.*, 19 (1954) 62-66.
- 11 G. W. ANDERSON, J. BLODINGER, AND A. D. WELCHER, *J. Amer. Chem. Soc.*, 74 (1952) 5309-5312.
- 12 J. E. SHIELDS, W. H. MCGREGOR, AND F. M. CARPENTER, *J. Org. Chem.*, 26 (1961) 1491-1494.
- 13 E. SCHROEDER AND E. KLIEGER, *Ann.*, 673 (1964) 208-220.
- 14 E. BELLEAU AND G. MALEK, *J. Amer. Chem. Soc.*, 90 (1968) 1651-1652.